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(54) Title: DETECTION PROBES, KITS AND ASSAYS			
(57) Abstract			
Nucleic acid hybridization probes having a first conformation when not interacting with a target and a second conformation when interacting with a target, and having the ability to bring a label pair into touching contact in one conformation but not the other, are labeled with a non-FRET pair of chromophores and generate a fluorescent or absorbance signal. Kits may include such probes, and assays, including multiplex assays, may utilize such probes.			

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## DETECTION PROBES, KITS AND ASSAYS

This invention was in part made with government support under grant number HL-43521-07, awarded by the National Institutes of Health. The United States 5 government has certain rights in that part of the invention.

This invention relates to nucleic acid hybridization probes containing fluorophore and chromophore labels, and kits and assays containing and 10 employing them.

### BACKGROUND

There are many different types of assays that employ nucleic acid hybridization probes and that utilize for signal generation a change in the fluorescence of a 15 fluorophore due to a change in its interaction with another molecule or moiety brought about by changing the distance between the fluorophore and the interacting molecule or moiety.

These assays rely for signal generation on 20 fluorescence resonance energy transfer, or "FRET", according to which a change in fluorescence is caused by a change in the distance separating a first fluorophore from an interacting resonance energy acceptor, either another fluorophore or a quencher. Combinations of a 25 fluorophore and an interacting molecule or moiety, including quenching molecules or moieties, are known as "FRET pairs." The mechanism of FRET-pair interaction requires that the absorption spectrum of one member of the pair overlaps the emission spectrum of the other 30 member, the first fluorophore. If the interacting molecule or moiety is a quencher, its absorption spectrum must overlap the emission spectrum of the fluorophore. Stryer, L., "Fluorescence Energy Transfer as a

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Spectroscopic Ruler," Ann. Rev. Biochem. 1978, 47: 819-846 ("Stryer, L. 1978"); BIOPHYSICAL CHEMISTRY part II, Techniques for the Study of Biological Structure and Function, C.R. Cantor and P.R. Schimmel, pages 448-455  
5 (W.H. Freeman and Co., San Francisco, U.S.A., 1980) ("Cantor and Schimmel 1980"), and Selvin, P.R., "Fluorescence Resonance Energy Transfer," Methods in Enzymology 246: 300-335 (1995) ("Selvin, P.R. 1995"). Efficient, or a substantial degree of, FRET interaction  
10 requires that the absorption and emission spectra of the pair have a large degree of overlap. The efficiency of FRET interaction is linearly proportional to that overlap. Haugland, R.P., Yguerabide, Jr., and Stryer, L., "Dependence of the Kinetics of Singlet-Singlet Energy  
15 Transfer on Spectral Overlap," P.N.A.S. (U.S.A.) 63: 24-30 (1969) ("Haugland et al. 1969"). The cited art teaches that to obtain a large magnitude of signal, a high degree of overlap is required. FRET pairs, including fluorophore-quencher pairs, have been chosen on  
20 that basis.

One suitable FRET pair disclosed in Matayoshi et al. 1990, Science 247: 954-958, includes DABCYL as a quenching moiety (or quenching label) and EDANS as a fluorophore (or fluorescent label). The absorption  
25 spectrum of DABCYL has a high degree of overlap with the emission spectrum of EDANS, making these two a good FRET pair. Despite the recognized advantage of using a FRET pair that includes a quencher that does not itself fluoresce, such as DABCYL, very few such FRET pairs have  
30 been identified. In general, the number of fluorophore-quencher pairs is extremely limited because of the need for a high degree of spectral overlap. Additional such pairs would be highly desirable for a number of reasons, including flexibility in assay design, distinguishing  
35 signals in multiplex assays.

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A variety of labeled nucleic acid hybridization probes and detection assays that utilize FRET and FRET pairs are known. One such scheme is described by Cardullo et al. (1988), P.N.A.S. 85: 8790-8794 and in 5 Heller et al. EP 0070685.A2, claiming priority of U.S. 284469 filed on July 17, 1981. It uses a probe comprising a pair of oligodeoxynucleotides complementary to contiguous regions of a target DNA strand. One probe molecule contains a fluorescent label, a fluorophore, on 10 its 5' end, and the other probe molecule contains a different fluorescent label, also a fluorophore, on its 3' end. When the probe is hybridized to the target sequence, the two labels are brought very close to each other. When the sample is stimulated by light of an 15 appropriate frequency, fluorescence resonance energy transfer from one label to the other occurs. FRET produces a measurable change in spectral response from the labels, signaling the presence of targets. One label could be a "quencher," which in this application is meant 20 an interactive moiety (or molecule) that releases the accepted energy as heat.

Another solution-phase scheme utilizes a probe comprising a pair of oligodeoxynucleotides and a FRET pair. However, here the two probe molecules are 25 completely complementary both to each other and to complementary strands of a target DNA (Morrison and Stols, "Sensitive Fluorescence-Based Thermodynamic and Kinetic Measurements of DNA Hybridization in Solution," Biochemistry 32: 309-3104 (1993) and Morrison EP 0 232 30 967 A2, claiming priority of U.S. application Serial No. 817841, filed January 10, 1986. Each probe molecule includes a fluorophore conjugated to its 3' end and a quenching moiety conjugated to its 5' end. When the two oligonucleotide probe molecules are annealed to each 35 other, the fluorophore of each is held in close proximity

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to the quenching moiety of the other. With the probe in this conformation, if the fluorophore is then stimulated by light of an appropriate wavelength, the fluorescence is quenched by the quenching moiety. However, when 5 either probe molecule is bound to a target, the quenching effect of the complementary probe molecule is absent. In this conformation a signal is generated. The probe molecules are too long to self-quench by FRET when in the target-bound conformation.

10 A solution-phase scheme that utilizes FRET pairs and the phenomenon known as strand displacement is described by Diamond et al. U.S. Patent No. 4,766,062; Collins et al. U.S. Patent No. 4,752,566; Fritsch et al. U.S. Patent Nos. 4,725,536 and 4,725,537. Typically, 15 these assays involve a probe comprising a bimolecular nucleic acid complex. A shorter single strand comprising a subset of the target sequence is annealed to a longer single strand which comprises the entire target binding region of the probe. The probe in this configuration thus comprises both single-stranded and double-stranded 20 portions. Diamond et al. proposed that these probes may further comprise either a  $^{32}\text{P}$  label attached to the shorter strand or a fluorophore and a quencher moiety which could be held in proximity to each other when the 25 probe conformation is that complex.

Another type of molecular probe assay utilizing a FRET pair is described in European Patent Application 0 601 889 A3, publication date 15 June, 1994, which claims priority of Bagwell U.S. patent application Serial No. 30 990,298, filed 10 December 1992.

Another type of nucleic acid hybridization probe assay utilizing a FRET pair is the so-called "TaqMan" assay described in Gelfand et al. U.S. Patent 5,210,015, and Livak et al. U.S. Patent 5,538,848. The 35 probe is a single-stranded oligonucleotide labeled with a

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FRET pair. In a "TaqMan" assay, a DNA polymerase releases single or multiple nucleotides by cleavage of the oligonucleotide probe when it is hybridized to a target strand. That release provides a way to separate 5 the quencher label and the fluorophore label of the FRET pair. According to Livak et al. "straightening" of an end-labelled "TaqMan" probe also reduces quenching.

Yet another type of nucleic acid hybridization probe assay utilizing FRET pairs is described in Tyagi et 10 al. copending United States patent application Serial No. 08/439,819 (and counterpart applications outside the United States, including PCT Application No. WO 95/13399), which utilizes labeled oligonucleotide probes, which we have come to refer to as "Molecular Beacons." 15 Tyagi, S. and Kramer, F.R., "Molecular Beacons: Probes that Fluoresce upon Hybridization," Nature Biotechnology 14: 303-308 (1996). A Molecular beacon probe is an oligonucleotide whose end regions hybridize with one another in the absence of target but are separated if the 20 central portion of the probe hybridizes to its target sequence. The rigidity of the probe-target hybrid precludes the simultaneous existence of both the probe-target hybrid and the intramolecular hybrid formed by the end regions. Consequently, the probe undergoes a 25 conformational change in which the smaller hybrid formed by the end regions disassociates, and the end regions are separated from each other by the rigid probe-target hybrid.

Aspects of this invention include probes 30 containing non-FRET fluorophore-quencher pairs and chromophore pairs useful in assays; improved assays, including multiplexed assays, utilizing such pairs of molecules or moieties; and assay kits that include such pairs.

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SUMMARY OF THE INVENTION

As opposed to FRET, quenching molecules and even other fluorophores can serve as efficient quenching moieties for fluorophores when attached to nucleic acid hybridization probes such that the fluorescing moiety and quenching moiety are in contact, even when the rules of FRET are violated. Further, the absorption spectra of a pair of chromophores (fluorescing or non-fluorescing), even identical chromophores, in a probe so constructed is altered in a detectable fashion.

In FRET, a first fluorophore absorbs at a first wavelength and emits at a second, longer wavelength. A second fluorophore or quencher which is near the first (the FRET range is reportedly 10-100 Å), if and to the degree its absorption spectrum overlaps that emission, absorbs some or most of the emitted energy and, if a fluorophore, re-emits at a third, still longer wavelength, or, if a quencher, releases the energy as heat. FRET progresses in the direction of increasing wavelength. It does not progress in the direction of decreasing wavelength. A probe according to this invention is a "nucleic acid hybridization probe," which as used herein means a probe that is hybridizable to a natural oligonucleotide strand and is composed of natural or modified nucleotides joined by natural or non-natural linkages. A peptide nucleic acid probe is a "nucleic acid hybridization probe," as are, of course, DNA probes, RNA probes and probes of mixed DNA and RNA. The term "probe" as used herein includes a single molecule and also a pair of molecules that in combination affect the level of signal.

A probe according to this invention is capable of undergoing a conformational change upon interacting with a target in an assay. Preferably a pair of labels, for example, a fluorophore and a quencher, "touch" all or

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part of the time when the probe is not interacting with target, and interaction with target separates the label pair, thereby preventing "touching". Examples of such probes include certain of the following probe

5 constructions: bimolecular probes disclosed in Morrison EP 0 232 967 A2, bimolecular probes disclosed in Diamond et al. U.S. Patent No. 4,766,062, single-molecule oligonucleotide probes such as disclosed in Gelfand et al. U.S. Patent 5,210,015 and Livak et al. U.S. Patent 10 No. 5,538,848 ("TaqMan" probes) and self-hybridizing single-molecule probes disclosed in Tyagi et al. U.S. Patent Application Serial No. 08/439,819, PCT Application No. WO 95/13399 and Nature Biotechnology 14: 303-308 (1996) ("Molecular Beacon" probes). However, a probe 15 according to this invention may function in the opposite manner such that a label pair is made to "touch" by interaction of the probe with its target. An example of such a probe is the bimolecular probe disclosed in Heller et al. EP 0070685.A2. Our most preferred probe 20 constructions are "Molecular Beacon" probes.

When attached to a probe according to this invention such that it is in contact with, or "touching", the first fluorophore in one of its conformations, a quenching moiety need not have an absorption spectrum 25 that overlaps the emission spectrum of the first fluorophore. Moreover, the absorption wavelength of the quencher can be shorter than the fluorophore's emission wavelength. Similarly, a second fluorophore that absorbs at a wavelength shorter than the emission wavelength of 30 the first can, in the probe construction described above, act as a quencher; that is, suppress emission by the first fluorophore and dissipate the incident energy as heat rather than as photon emission.

In probes constructed as described above, 35 change in the absorption spectra of the label pair can be

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used as a detectable signal, as an alternative to change in fluorescence. When change in absorption is utilized, the label pair may include any two chromophores, that is, fluorophores, quenchers and other chromophores. The 5 label pair may even be identical chromophores.

In addition to probes, this invention includes assays employing such probes and assay kits containing them.

BRIEF DESCRIPTION OF THE DRAWINGS

10 Figure 1 shows excitation and emission spectra for EDANS-labeled DNA.

Figure 2 shows excitation and emission spectra for Tetramethylrhodamine-labeled DNA.

15 Figure 3 is a graph showing the absorption spectrum of DABCYL and emission spectra for each of nine fluorophores.

Figure 4 shows the chemical structure of reactive forms of a number of fluorophores.

20 Figure 5 shows the chemical structures of reactive forms of several quenchers and a fluorophore used as a quencher.

Figure 6 shows absorption spectra for a pair of chromophores when touching and when separated.

25 Figure 7 shows kinetic fluorescence measurements using four probes in a multiplex assay.

Figure 8 shows real-time fluorescence measurements using two probes in a multiplex PCR amplification assay.

DETAILED DESCRIPTION OF THE INVENTION

30 One type of probe structure useful in this invention is the "Molecular Beacon" oligonucleotide

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structure described in U.S. Patent Application Serial No. 08/439,819, incorporated by reference herein in its entirety, PCT Application No. WO 95/13399, and Tyagi, S. and Kramer, F.R., "Molecular Beacons: Probes that 5 Fluoresce upon Hybridization," Nature Biotechnology 14: 303-308 (1996). In those probes, a central target-recognition sequence is flanked by arms that hybridize to one another when the probe is not hybridized to a target strand, forming a "hairpin" structure, in which the 10 target-recognition sequence (which is commonly referred to as the "probe sequence") is in the single-stranded loop of the hairpin structure, and the arm sequences form a double-stranded stem hybrid. When the probe hybridizes to a target, that is, when the target-recognition 15 sequence hybridizes to a complementary target sequence, a relatively rigid helix is formed, causing the stem hybrid to unwind and forcing the arms apart. A FRET pair, such as the fluorophore EDANS and the quencher DABCYL, may be attached to the arms by alkyl spacers. When the 20 Molecular Beacon is not hybridized to a target strand, the fluorophore's emission is quenched. When the Molecular Beacon is hybridized to a target strand, the FRET pair is separated by more than 100 Å, and the fluorophore's emission is not quenched. Emitted 25 fluorescence signals the presence of target strands.

Molecular beacon probes may have target recognition sequences 7-140 nucleotides in length and arms that form a stem hybrid, or "stem duplex" 3-25 nucleotides in length. Modified nucleotides and modified nucleotide 30 linkages may be used. Molecular beacons may be, for example, peptide nucleic acid ("PNA") probes.

We have discovered that in certain Molecular Beacon probes, a pair of labels "touches" when the probe is not hybridized to a target. By "touching" we mean 35 that the absorption spectrum of the pair is significantly

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altered. Figure 6 presents absorption spectra for a Molecular Beacon whose complementary terminal nucleotides are part of a stem hybrid. The probe was labeled with DABCYL on one end and Tetramethylrhodamine on the other 5 end. Spectrum 20 is from the probe in a first conformation, not bound to target, with the arms hybridized to one another. Spectrum 21 is from the probe in a second conformation, bound to target, with the arms not hybridized. The absorption maxima are at about 4800 10 Å and 5500 Å in spectrum 21. Comparison of spectrum 20 with spectrum 21 shows that the label pair are "touching" when the arms are hybridized: absorption at 4800 Å is reduced from about 0.033 to about 0.026, and absorption at 5500 Å is increased from about 0.046 to about 0.063. 15 FRET would not change the absorption spectrum of the pair. Van Der Meer, B.W., Coker, G., III, and Chen S-Y.S., RESONANCE ENERGY TRANSFER, VCH Publishers, Inc. (New York 1994). Whether or not two chromophores are "touching" can be determined by comparing the absorption 20 spectrum when they are touching to the absorption spectrum when they are separated by more than 100 Å, as shown in Figure 6.

We have discovered that a quencher, for example DABCYL, attached to one end of a Molecular Beacon 25 constructed as described in the preceding paragraph, will effectively quench fluorophores attached to the other end in violation of FRET rules, thereby greatly enlarging the available number of fluorophore-quencher pairs that can be used in nucleic acid hybridization probes.

30 The following description references the spectral data presented in Figures 1, 2 and 3. Figure 1 presents the excitation (curve 1 on the left, shorter wavelength) and emission (curve 2 on the right, longer wavelength) spectra of a Molecular Beacon containing the 35 quencher DABCYL and the fluorophore EDANS, when it is

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hybridized to a target. Figure 2 presents the excitation spectrum (curve 3) and emission spectrum (curve 4) of a Molecular Beacon containing the quencher DABCYL and the fluorophore Tetramethylrhodamine, when it is hybridized 5 to a target.

Figure 3 presents the absorption spectrum 5 for the quencher DABCYL and emission spectra 6-14 for a number of fluorophores: EDANS(6), Fluorescein(7), Lucifer Yellow(8), BODIPY(9), Eosine(10), Erythrosine(11), 10 Tetramethylrhodamine(12), Texas Red(13) and Coumarin(14). The large overlap for EDANS and Coumarin with DABCYL demonstrates why EDANS and DABCYL and Coumarin and DABCYL are FRET pairs. Curve 12 is the emission spectrum for Tetramethylrhodamine. It has almost no overlap with the 15 absorbance spectrum of DABCYL. Curve 13 is the emission spectrum for Texas Red. It has no overlap with the absorbance spectrum of DABCYL. According to FRET rules, the degree of fluorescence energy transfer is a function of the amount of spectral overlap.

20 Initially using the procedure described in Example 1, we determined the spectral overlap of several fluorophores with an exemplary quencher, for which we chose DABCYL. The calculated overlap and the consequent expected degree of quenching are summarized in Table 1.

25

Table 1

Quenching with DABCYL by FRET

<u>Fluorophore</u>	<u>Spectral Overlap</u>	<u>Expected FRET</u>	<u>Efficiency, %</u>
	<u>Quenching</u>		
EDANS	4856		95.60
Fluorescein	1448		30.38
Lucifer Yellow	2485		52.15
BODIPY	1050		22.04

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Eosine	409	8.58
Erythrosine	184	3.86
Tetramethylrhodamine	2	0.05
Texas Red	0	0.00

5           For use in an assay, a quencher or interacting  
fluorophore should have sufficient spectral overlap, as  
spectral overlap was determined by the procedure of  
Example 1, to absorb at least 60% of a fluorophore's  
emission by fluorescence resonance energy transfer, which  
10 we define as the minimal interaction to be considered a  
"FRET pair" as that term is used herein. According to  
that description, only EDANS of the fluorophores in Table  
1 forms a FRET pair with the quencher DABCYL. We have  
discovered, however, that when DABCYL or another suitable  
15 quencher contacts or "touches" any of the seven other  
fluorophores tested, efficient quenching is achieved.

          To demonstrate embodiments of probes with  
"touching" pairs of a fluorophore with another  
fluorophore or quencher, where the pairs are not FRET  
20 pairs as defined above, we prepared Molecular Beacon  
probes end-labeled with DABCYL at one end and one of  
eight different fluorophores at the other end. We tested  
quenching efficiency by the procedures described in  
Example 2. Table 2 presents the observed quenching  
25 efficiency and also the expected quenching efficiency by

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FRET. Figure 6 demonstrates that "touching" is achieved, and Table 2 shows the effect on quenching that results for non-FRET pairs, which includes all fluorophores in Table 2 except EDANS.

5

Table 2

Quenching by "Touching"

	<u>Fluorophore</u>	<u>Expected FRET Quenching</u>	<u>Observed</u>
	<u>Quenching</u>	<u>Efficiency, %</u>	<u>Efficiency, %</u>
10	EDANS	95.60	95.60
	Fluorescein	30.38	94.52
	Lucifer Yellow	52.15	96.84
	BODIPY	22.04	94.17
15	Eosine	8.58	90.66
	Erythrosine	3.86	72.22
	Tetramethylrhodamine	0.05	95.77
	Texas Red	0.00	99.10

Not all Molecular Beacon designs are considered  
20 likely candidates for probes according to this invention,  
because "touching" is required. Only Molecular Beacons  
having a label pair attached such that the label moieties  
can touch when the probe is not hybridized to a target  
may be probes according to this invention. Besides the  
25 end-labeling described above, another configuration that

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should be useful is to separate the label moieties by five nucleotides along the stem hybrid. This brings the two labels to the same side of the helix, because the helix makes a complete turn every ten nucleotides.

5 Separation by four or six nucleotides along the helix would be less preferred candidates. To ascertain whether or not a particular Molecular Beacon design is suitable for probes according to this invention, one can simply prepare a probe and test it by the procedures described  
10 herein.

The same applies to other probe constructions. The data presented here establishes that probe pairs of the types described in Heller et al. EP 0070685.A2, Morrison EP 0232967.A2 and Diamond U.S. 4,766,062 will  
15 provide "touching" if the two molecules of the probe are end-labeled in the fashion described herein for Molecular Beacons. Other candidate configurations should be tested.

Probes according to this invention include  
20 probes whose labels can "touch", but in fact "touch" only part of the time, such as, for example, a "TaqMan" probe or other single-stranded probe that is labeled along the probe. Such linear probes form a random coil when not hybridized. Probes that "touch" only part of the time  
25 will tend to be less efficiently quenched and generate higher background than probes such as the preferred

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Molecular Beacon probes described herein and are, therefore, less preferred. To be useful in this invention such a probe must provide at least 20% quenching, preferably 30% and more preferably at least 5 40% quenching, according to the procedure of Example 2.

To demonstrate probes whose labels "touch" only part of the time, we prepared three non-self-hybridizing oligonucleotide probes. They were thirteen nucleotides long and end-labeled with DABCYL (3' end) and one of 10 three fluorophores -- EDANS, Fluorescein or Tetramethylrhodamine (5' end). We hybridized the three probes to an unlabeled, perfectly complementary target oligonucleotide, of the same length as the probes and we hybridized the three probes to a perfectly matched target 15 oligonucleotide labeled at its 3' end with DABCYL. The first target eliminates quenching, and the second target maximizes quenching by "touching" the fluorophore to the target's DABCYL moiety. Utilizing the procedure of Example 2, we measured quenching efficiency when the 20 probes were not hybridized (a random-coil state) and hybridized to each target. Table 3 presents those results.

Table 3

Observed Quenching Efficiency, %

25 Probe/State                   fluorophore paired with DABCYL

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EDANS Fluorescein Tetramethyl-  
rhodamine

		EDANS	Fluorescein	Tetramethyl-
				rhodamine
	13-mer/unhybridized	76	81	44
5	13-mer/hybridized to unlabeled target	0	0	0
	13-mer/hybridized to labeled target	96	95	96

The results in Table 3 show that for a probe according to this invention in which the label pair 10 (Tetramethylrhodamine-DABCYL) has essentially no spectral overlap, precluding FRET interaction completely, nearly half of the fluorescence (44%) was quenched by "touching" part of the time due to random-coil movement. A probe according to this invention in which the label pair 15 (Fluorescein-DABCYL) has some spectral overlap but not enough to be a FRET pair was quenched as well as the probe with the FRET pair (EDANS-DABCYL). The background from all three 13-mer oligonucleotide probes was shown to be considerably higher than from similarly labeled 20 Molecular Beacon probes, which give better quenching and, thus, higher signal-to-background ratios.

To be useful in this invention, a non-FRET fluorophore-quencher pair when attached to complementary ends of a hybrid must satisfy certain criteria when 25 tested according to Examples 1 and 2 presented below. The pair must yield a degree of quenching above 60 percent, preferably at least 70 percent and more

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preferably at least 80% to minimize background signal.

In addition, the quenching efficiency must be at least 10 percent greater than predicted by spectral overlap, which follows from the significant change in absorbance that

5 signifies "touching". Preferably, the quenching efficiency is at least 30 percent greater than predicted by spectral overlap.

A quencher suitable for use in this invention should satisfy several criteria. First, it is a  
10 chromophore that is not a fluorophore; it can absorb light and dissipate energy as heat. Second, it should be a relatively good absorber; it preferably should have an extinction coefficient of at least  $10^4 \text{ M}^{-1}$ . Third, it should not repel or seek to avoid the fluorophore to be  
15 quenched; that is, if the fluorophore is hydrophobic, the quencher should also be hydrophobic, and if the fluorophore is positively or negatively charged, the quencher should not be similarly charged. Fourth, it should not be chemically reactive in use, particularly it  
20 should not generate destructive free radicals. Finally, for ease of use, it should be available in an activated form containing leaving groups for coupling to the probe. Quenchers satisfying these criteria include, for example,  
25 DABCYL, DABMI and Malachite Green, all of which appear in Figure 5.

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A second fluorophore that does not form a FRET pair with the first fluorophore can be used in place of a quencher. Preferably the second fluorophore has no spectral overlap or minimal spectral overlap with the 5 first fluorophore. Criteria for selecting a second fluorophore are basically the same as for selecting a quencher. Considering the first fluorophore to be the one excited in an assay, it is preferred that the excitation spectrum of the second fluorophore be at 10 wavelengths shorter, completely or nearly so, than the emission excitation spectrum of the first fluorophore. Coumarin, a fluorophore that normally emits in the blue range, has been shown to act as a quencher for Tetramethylrhodamine in a Molecular Beacon probe 15 according to this invention.

Although change in fluorescence is preferred for detection in this invention, "touching" permits change in absorbance to be used for detection. As explained above in connection with Figure 6, the 20 absorption spectrum changes when two chromophores, fluorescing or non-fluorescing, are made to "touch". We have shown this occurs, utilizing a Molecular Beacon labeled at both ends with Tetramethylrhodamine. Other label pairs could be, for example, the non-fluorophores 25 DABCYL-DABCYL, Malachite Green-DABCYL or Malachite Green-Malachite Green, or the fluorophores Fluorescein-

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Tetramethylrhodamine. Figure 6 demonstrates that the label pair could be DABCYL-Tetramethylrhodamine. For use with absorption detection, one could select a suitable pair by obtaining absorption spectra for the probe in the 5 "touching" and "non-touching" configurations, and ascertaining if the change in absorption is a suitable signal for the test or assay and the sensitivity of the instrument to be used. Where two different chromophores are used, the change in the ratio of absorbencies at 10 their peak wavelengths is a convenient way to measure signal change. Preferably, where absorbance is used for detection, the labels should have extinction coefficients of at least  $3 \times 10^4 \text{ M}^{-1}$ .

Attachment of chromophores to the probe is by 15 conventional means. The points of attachment, taking into consideration linkers, such as alkyl spacers, for example, must permit the two labels to "touch" as defined above. Complementary end nucleotides of a hybrid (whether in one or separate molecules) ensure "touching", 20 as demonstrated by the Molecular Beacons described herein.

This invention also includes assays that utilize chromophore pairs, preferably fluorescer-quencher pairs, of this invention. Thus, in an assay that relies 25 for generation of a signal on quenching the fluorescence of a fluorophore, this invention includes the improvement

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comprising using a fluorophore-quencher pair wherein the quencher is, for example, DABCYL and the degree of quenching is at least 60%, preferably 70% and more preferably 80%, and at least 10%, preferably at least 5 30%, above the quenching efficiency expected based on spectral overlap. An assay according to this invention can be a multiplex assay that includes multiple fluorescer-quencher pairs and generates multiple signals. The assays may be detection assays, either qualitative, 10 quantitative or both. Detection may comprise part of a larger process that includes, for example, a nucleic acid amplification. This will be the case particularly if detection is in real-time during the amplification or other process. Well-known amplification processes 15 include the polymerization chain reaction ("PCR") process disclosed, for example, in United States Patent Nos. 4,683,202, 4,683,195 and 4,965,188; Q-beta replication (Lomeli et al., "Quantitative Assays Based on the Use of Replicable Hybridization Probes," Clin. Chem. 39, 1826- 20 1831 (1989)); NASBA; self-sustained sequence reactions ("3SR") (Guatelli et al., "Isothermal in vitro Amplification of Nucleic Acids by Multienzyme Reaction Modeled after Retroviral Replication," P.N.A.S. (U.S.A.) 87: 1874-1878 (1990)) and transcription and replication 25 reactions.

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Our invention also includes reagent kits that include labeled probes according to this invention, together with other reagents for an assay. For example, a kit may include enzymes, primers and buffers for a PCR reaction together with Molecular Beacons for detecting amplified product. For multiplex assays, kits according to this invention will include multiple probes, at least one of which is a probe according to this invention. For example, several Molecular Beacons can be included, each 10 labeled according to this invention with DABCYL as a quencher, but each with a different fluorophore. In that case at least one probe will include a non-FRET fluorophore-quencher pair according to this invention. Another pair may be a recognized FRET pair, such as EDANS 15 and DABCYL. Multiple probes according to this invention may, of course, be included. For multiplexing with fluorescence detection, it will be recognized that it is desirable to minimize the overlap of both the excitation spectra and the emission spectra of the different 20 fluorophores. For multiplexing with absorption detection, it will be recognized that it is desirable to minimize the overlap of absorption spectra of the different probes.

Using the assay procedures described herein, we 25 have shown that DABCYL can quench the fluorescence of Fluorescein, Lucifer Yellow, BODIPY, Eosine, Erythrosine,

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Tetramethylrhodamine, Texas Red and Coumarin, all commercially available fluorophores whose chemical structures are well known and published. We have also shown effective fluorophore quenching in non-FRET pairs 5 containing other quenchers, DABMI and Malachite Green, as well as appropriate (shorter wavelength) fluorophores such as Coumarin. Figure 4 shows the chemical structures of the reactive forms of fluorophores, and Figure 5 shows the chemical structures of the reactive forms of 10 Coumarin, DABMI, Malachite Green and DABCYL, as reported in the catalog of Molecular Probes, Inc., Eugene, Oregon, U.S.A., entitled "Handbook of Fluorescent Probes and Research Chemicals," 5th Ed. 1992-1994, by R.P. Haugland (K.D. Larison, ed.), copyright 1992. These combinations 15 are given by way of example only. Workers in the art can, by the straightforward procedures set forth herein, select and test other chromophores.

Example 1

Initial Determination of Spectral Overlap

20 In order to determine the spectral overlap between the absorption spectrum of DABCYL and emission spectrum of each of several fluorophores, the absorption spectrum of DABCYL was determined. Since the absorption spectra of DABCYL may be altered upon attachment with 25 DNA, we coupled DABCYL to an oligodeoxynucleotide by

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using methods described by Tyagi and Kramer 1996. The oligodeoxynucleotide was 5' AAG TTA AGA CCT ATG A - hexalkylamino-DABCYL. A  $6 \times 10^{-6}$  solution (400  $\mu$ ) of this oligodeoxynucleotide was placed in a quartz cuvette 5 and its absorption spectra was determined using a Perkin-Elmer Lambda 3A UV-Visible spectrophotometer. The visible portion of this spectrum is shown in Figure 3. The emission spectra of molecular beacons containing different fluorophores (each bound to their targets) is 10 also plotted in the same figure. Each spectrum was normalized by assigning a value of 100 to its maxima and a proportionally lower value to other points. For each emission spectrum, only the top 75% of the spectrum on either side of the maxima were considered. By "top 75%" 15 is meant the range of emission wavelength for which the emission is at least 75% of the maximum emission. The extent of overlap between the absorption spectrum of DABCYL and the emission spectrum of various fluorophore was determined graphically. The spectral overlap for 20 EDANS was, sum of absorption of DABCYL at each wavelength between 490 nm and 524 nm plus sum of emission of EDANS at each wavelength from 469 nm to 489 nm. For Fluorescein it was, sum of absorption of DABCYL at each wavelength between 507 nm and 527 nm plus sum of emission 25 of Fluorescein at each wavelength from 505 nm to 530 nm. For Lucifer Yellow it was the sum of absorption of DABCYL

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at each wavelength between 506 nm and 548 nm plus the sum of emission of Lucifer Yellow at each wavelength from 520 nm to 504 nm. For BODIPY, Eosine, Erythrosine, and Tetramethylrhodamine the overlap was the sum of DABCYL absorption from 513 nm to 533 nm, 532 nm to 553 nm, 542 nm to 563nm, and 560 nm to 588 nm respectively. For Texas Red shown in Figure 3) there was no overlap at all, even when the entire emission spectrum was considered. The extents of spectral overlaps, which according to our definition has no units, are shown in Table 1.

Calculation Expected Quenching Efficiencies

For a chemical moiety to serve as a quencher of a fluorophore by FRET, its absorption spectrum must overlap with the emission spectrum of the fluorophore (Stryer, L. 1978; Cantor and Schimmel 1980; and Selvin P.R. 1995). The efficiency of FRET (and therefore quenching) between a fluorophore and a quencher is linearly proportional to the spectral overlap between the absorption spectrum of the quencher and the emission spectrum of the fluorophore. This prediction from Forster's original theory of fluorescence resonance energy transfer was confirmed experimentally (Haugland et al. 1969) and has since been the basis of selection of FRET pairs. The expected efficiency or degree of quenching for the set of fluorophores, assuming a linear

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relationship between the quenching efficiency and the spectral overlap, are shown in Table 1. In order to calculate the expected efficiency or degree of quenching, the spectral overlap for each fluorophore was divided by 5 the spectral overlap for EDANS and multiplied by the observed quenching efficiency of EDANS. The expected degrees of quenching are also presented in Table 1.

Example 2

10 Synthesis of Molecular Beacons to Test the Degree of Quenching of Candidate Fluorophores by DABCYL

We prepared eight molecular beacons with identical nucleotide sequences. Each of these molecular beacon contained a DABCYL moiety at its 3' end and one of six candidate fluorophores that we tested (Fluorescein, 15 BODIPY, Lucifer Yellow, EDANS, Erythrosine, Eosine, Tetramethylrhodamine or Texas Red) at its 5' end. Thus they were identical except the nature of fluorophore that was present at their 5' end. In order to synthesize these molecules, an oligonucleotide which contained a 20 protected sulphydryl group at its 5' end and a primary amino group at its 3' end was obtained from Midland Certified Reagents (Midland Texas). The sulphydryl and the primary amino group in this oligonucleotide were tethered to the DNA via - (CH<sub>2</sub>)<sub>6</sub> - and - (CH<sub>2</sub>)<sub>7</sub> - spacers 25 respectively. The sequence of the oligonucleotide was: 5' SH - CCG AGA AAG AAA ATA TCA TTG GTG TTT CCT ATG ATG

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AAT CTC GG - Amino 3'. An amine reactive derivative of DABCYL was coupled to the 3' end of this oligonucleotide according to the methods described by Tyagi and Kramer 1996. The coupled oligonucleotide was purified using 5 high pressure liquid chromatography. After purification, the protective group was removed from the sulphydryl group at the 5' end (Tyagi and Kramer 1996). This preparation was divided into seven fractions. These fractions were coupled with iodoacetamide derivatives of 10 Fluorescein, Lucifer Yellow, EDANS, Erythrosine, Eosine and Tetramethylrhodamine and with the bromomethyl derivative of BODIPY (Molecular Probes). In each case, the unreacted fluorophore was removed by gel filtration on a small column (NP-5, Pharmacia). Each doubly labeled 15 oligonucleotide was further purified by high pressure liquid chromatography and dissolved in buffer A (1 mM MgCl<sub>2</sub>, and 20 mM Tris-HCl pH 8.0).

#### Determination of Quenching Efficiency

The molecular beacons labelled with DABCYL at 20 their 3' ends and a fluorophore at their 5' ends were designed in such a way that six terminal nucleotides on either side of the molecule formed a hairpin stem which brought the fluorophore in close proximity to the DABCYL. With a FRET pair, it was expected from Tyagi et al. that 25 quenching would occur by fluorescence resonance energy

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transfer. With non-FRET pairs, as defined herein, no quenching or less than minimally needed quenching was expected by fluorescence resonance energy transfer.

The middle region of these oligonucleotides 5 constitutes a probe that can interact with a target. The interaction of the probe with a target leads to unwinding of the stem helix and separation of the fluorophore from the DABCYL by more than 100 Å. In this state, no fluorescence resonance energy transfer can occur and 10 therefore no quenching can take place. Tyagi and Kramer have shown that when EDANS and DABCYL are used as a fluorophore and quencher of a FRET pair in these oligonucleotides, the fluorescence of EDANS is quenched very efficiently by DABCYL. Upon addition of the target 15 of the molecular beacon the fluorescence of EDANS is fully restored.

In order to determine the degree of quenching for the present series of fluorophores, the excitation and emission spectra of each molecular beacon was 20 recorded before and after addition of the target using a Photon Technology (Princeton) fluorometer. A solution of a molecular beacon in Buffer A was held at 25°C in a cuvette. The solution was excited at the excitation maxima of the fluorophore in the molecular beacon, while 25 the intensity of the emitted light was measured as a function of emission wavelength. An excitation spectrum

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was recorded by monitoring the intensity of emission at the emission maximum of the fluorophore and changing the excitation wavelength. Sufficient time was given for the hybrids to form. After recording the two spectra, a  
5 five-fold molar excess of the target of the Molecular Beacon was added to the solution. After the completion of hybridization, the excitation and emission spectra were recorded again. Four spectra were recorded for each of the eight Molecular Beacons. The spectra for two,  
10 Molecular Beacons containing EDANS and Tetramethylrhodamine, are shown in Figures 1-2, where the spectra before the addition of the target (71, 72, 73, 74) are depicted by broken lines and the spectra after the addition of target (1, 2, 3, 4) are shown by  
15 continuous lines. The spectra on the left are excitation spectra. On the right are emission spectra.

The degree of quenching was determined from the intensities of emission at the emission maxima. The percentage of observed quenching is defined as  $(1 - \frac{F_o}{F_t}) * 100$ , where  $F_o$  is the intensity of emission before the addition of the target and  $F_t$  is the intensity of emission after the addition of the target. Table 2 lists the observed quenching efficiency for the seven fluorophores tested, as well as the expected quenching  
20 efficiency if FRET was the mechanism.  
25

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Example 3

In order to demonstrate the utility of fluorophore-quencher combinations that are available for use in probes of this invention, a multiplex detection assay utilizing four different nucleic acid targets was performed. We chose targets that differed in their nucleotide sequence, by just one nucleotide and designed a set of four differently labeled molecular beacons to detect these targets. These molecular beacons possessed a DABCYL moiety at their 3' end and were identical in all respects, except that each was labeled with a different fluorophore at its 5' end and each had a different nucleotide at the center of its probe sequence. These molecular beacons were labeled with either a blue, green, orange, or red fluorophore and had either a thymidine, cytidine, adenine, or guanosine residue at the center of its probe sequence, respectively. The nucleotide sequences of the blue, green, orange, and red molecular beacons were coumarin-5'-GCG AGC CAC CAA ATA TGA TAT GCT CGC-3'-DABCYL, Fluorescein-5'-GCG AGC CAC CAA ACA TGA TAT

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GCT CGC-3'-DABCYL, Tetramethylrhodamine-5'GCG AGC CAC CAA  
AAA TGA TAT GCT CGC-3'-DABCYL, and Texas red-5'-GCG AGC  
CAC CAA AGA TGA TAT GCT CGC-3'-DABCYL, respectively. The  
targets of these molecular beacons were 5'-AAA GAA AAT  
5 ATC ATAA TTT GGT GTT TCC TAT-3', 5'-AAA GAA AAT ATC ATG  
TTT GGT GTT TCC TAT-3', 5'-AAA GAA AAT ATC ATT TTT GGT  
GGT TCC TAT-3', 5'-AAA GAA AAT ATC ATC TTT GGT GTT TCC  
TAT-3', respectively. The underlined residues represent  
the only difference in the nucleotide sequence among  
10 these molecules. The fluorophores were chosen in such a  
way that when each was excited at its optimal excitation  
wavelength and observed at its optimal emission  
wavelength, the fluorescence of one molecular beacon  
could be determined independently of the fluorescence of  
15 each of other molecular beacons. An equimolar mixture of  
these molecular beacons was prepared and aliquots of this  
mixture were plated into each of four tubes. One of the  
four targets was added to each of these solutions while  
holding the temperature of the solution 25°C, and the  
20 fluorescence of each molecular beacon in that tube was  
monitored repeatedly. The changes in fluorescence of  
each molecular beacon was monitored by rapidly changing  
the position of the diffraction gratings of our  
spectrofluorometer to alternate between the optional pairs  
25 of excitation and emission wavelengths for measuring the  
fluorescence of each fluorophore. While holding the

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excitation wavelength at 400 nm, the fluorescence of Coumarin was monitored at 475 nm for a five second interval. These wavelengths were changed to 491 and 515 for the next five seconds (Fluorescein), and then to 575 5 and 595 for the following five seconds (Tetramethylrhodamine), and finally to 595 and 615 for the last five seconds (Texas Red). This series of measurements was made repeatedly until the fluorescence of the perfectly matched probe-target hybrid reached a 10 plateau. These results were plotted as, four separate fluorescence vs. time curves, one for each molecular beacon.

The addition of each target led to an increase in the fluorescence of the perfectly complementary 15 molecular beacon, but did not lead to an increase in the fluorescence of the other molecular beacons that were present in the same solution (Figure 7). When the target 100 contained an adenosine as its central nucleotide (Panel a), the blue fluorescence indicated by squares 101 increased, when the target 110 contained a guanosine as 20 its central nucleotide (Panel b), the green fluorescence indicated by circles 111 increased, when the target 120 contained a thymidine as its central nucleotide (Panel c), the orange fluorescence indicated by inverted 25 triangles 121 increased, and when the target 130 contained a cytidine as its central nucleotide (Panel d),

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the red fluorescence indicated by diamonds 131 increased. The color of the fluorescence in the tube indicated which nucleotide residue was present at the center of the target. There was no significant "crosstalk," either at 5 the level of hybridization, or at the level of signal detection. The two notable exceptions were that a target 110 with a guanosine residue at its center elicited a small response 112 from a molecular beacon that contained a thymidine at the corresponding position, and a target 10 120 with a thymidine at its center elicited a small response 122 from a molecular beacon that contained a guanosine at the corresponding position. This occurred because guanosine is able to form a relatively stable base-pair with thymidine. Each target could also be 15 identified visually by the color of the fluorescence that developed when higher concentrations of molecular beacons and targets were used and when the tubes were illuminated by a broad-wavelength ultraviolet lamp.

We also demonstrated the utility of this 20 invention in a very simple detection of the genetic abnormality that is responsible for cystic fibrosis. One of the most common genetic causes of cystic fibrosis is a three-nucleotide deletion at position 508 in the gene for the cystic fibrosis transmembrane conductance regulator 25 (CFTR). Two molecular beacons were prepared for the detection of this gene. One molecular beacon was

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labelled with Fluorescein and DABCYL and had a probe sequence that was perfectly complementary to the wild-type CFTR gene sequence. The other molecular beacon was labelled with Tetramethylrhodamine and DABCYL and had 5 a probe sequence that was perfectly complementary to the mutant CFTR gene sequenced. Three tubes containing an equimolar mixture of both of these molecular beacons and containing the other components necessary for a polymerase chain reaction (including appropriate 10 primers), were prepared. DNA encoding the wild-type gene was added to the first tube and DNA encoding the mutant gene was added to the second tube. The third tube was a negative control that did not contain any template DNA. Polymerase chain reactions were performed by cycling the 15 temperature of the tubes between 94, 55 and 72 degrees centigrade corresponding to the melting, annealing, and polymerization steps, respectively, of the PCR cycle. The progress of the reactions was monitored in the sealed tubes during the course of these reaction by measuring 20 the fluorescence of each molecular beacon repeatedly during the 55° cycle. A thermal cycler that could monitor the fluorescence of the sealed tubes in real-time was used for this purpose (System 7700, Applied Biosystems, CA). Figure 8 shows the results. The tube 25 that contained wild-type (template) DNA (Panel a) exhibited a rise in the fluorescence of the Fluorescein

fluorescein molecular beacon (squares 140), but did not exhibit a rise in the Tetramethylrhodamine fluorescence (circle 141). The tube that received mutant template DNA (Panel b) exhibited a rise in the fluorescence of 5 Tetramethylrhodamine (circles 150), but did not exhibit a rise in the fluorescence of fluorescein (151). The level of fluorescence of neither molecular beacon rose in the third tube. This background signal is indicated by inverted triangles (142,152 and is included in both of 10 the panels. This experiment demonstrates that by using the fluorophore-quencher pairs selected according to this invention, it is possible to perform multiplex detection of genetic alleles.

#### Example 4

Subsequent to our initial experiments on quenching according to the procedures set forth in Example 1 and Example 2, we carried out further 5 experiments with more extensively purified Molecular Beacon probes. Three HPLC purifications were used for en-labeled Molecular Beacon probes having a central region 20 nucleotides long flanked by arms 6 nucleotides long and completely complementary to each other. Label 10 pairs were DABCYL on one end and one of nine different fluorophores (Figure 3) on the other end.

Expected quenching efficiency was calculated differently from the method of Figure 2. Entire spectra were used as a basis, and calculation was according to 15 Haugland et al. 1969, discussed earlier, wherein spectral overlap is defined as,  $J = \int F(\lambda) \epsilon(\lambda) \lambda^4 d\lambda / \int F(\lambda) d\lambda$ , where,  $F(\lambda)$  is the normalized fluorescence emission at wavelength  $\lambda$ ; and  $\epsilon(\lambda)$  is the molar extinction coefficient of DABCYL when coupled to an oligonucleotide 20 at wavelength  $\lambda$ . All spectral parameters were determined in the buffer mentioned above using a photon counting fluorometer (Photon Technology International, NJ).

Utilizing full spectra, of course, the amount of overlap is greater than the amount calculated 25 according to Example 1, and, consequently, so is the expected quenching efficiency. Our definition of the minimum expected quenching efficiency required for a FRET pair and for a non-FRET pair useful in a probe according to this invention all change upwardly using this different technique. The minimum expected efficiency for 5 a FRET pair would be 80%, for example (rather than 60% according to the method of Example 1), and a probe according to this invention would have observed quenching efficiency of at least 80%, preferably 90% and more preferably 95%, plus be at least 10% above the expected

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10 from overlap, preferably at least 15%. The results obtained are presented in Table 4. To avoid confusion, the entirety of this specification and claims, except for this Example 4, is to be interpreted according to the definitions related to the procedures of Examples 1 and  
 15 2, not this Example 4.

Table 4

**Comparison of Expected and Observed  
Quenching Efficiency, %**

Fluorophore	Spectral Overlap ( $10^{-15} M^{-1} cm^{-3}$ )	Expected Quenching Efficiency, %	Observed Quenching Efficiency, %
Coumarin	1.260	99.3	99.3
EDANS	1.090	85.9	99.5
Fluorescein	0.974	76.7	99.9
Lucifer Yellow	0.796	62.7	99.2
BODIPY	0.781	60.9	95.0
Eosine	0.414	32.9	98.2
Erythrosine	0.253	19.9	87.5
Tetramethylrhodamine	0.019	1.4	98.7
5 Texas Red	0.000	0.0	99.1

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We claim:

1. A probe capable of hybridizing with a nucleic acid strand comprising one or two molecules from the group consisting of natural nucleotides, modified 5 nucleotides and combinations thereof, and a non-FRET pair of labels consisting of a first fluorophore and a chromophore, said chromophore selected from the group consisting of fluorophores and quenchers, wherein interaction of the probe with a target causes the probe 10 to change from a first conformation to a second conformation, thereby changing the distance between the labels of said label pair, and wherein in only one conformation do the labels touch sufficiently to quench 15 the fluorescence of said first fluorophore by at least 25 percent.

2. The probe according to claim 1 wherein said probe comprises two molecules that are capable of hybridizing with one another to form a hybrid and one member of said label pair is attached to one of said 20 molecules at an end of said hybrid, wherein the degree of quenching in the hybrid is at least sixty percent and at least ten percent above the degree of quenching expected based on spectral overlap.

3. The probe according to claim 1 wherein 25 said probe comprises a single molecule.

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4. The probe according to claim 3 wherein the members of said non-FRET label pair, when attached to hybridized end nucleotides of double stranded DNA, provide a degree of quenching of at least sixty percent 5 and at least ten percent above the degree of quenching expected based on spectral overlap.

5. The probe according to claim 4 wherein the members of said non-FRET label, when attached to hybridized end nucleotides of double stranded DNA, 10 provide a degree of quenching of at least seventy percent and at least thirty percent above the degree of quenching expected based on spectral overlap.

6. The probe of claim 4 or claim 5 wherein the chromophore is a quencher.

15 7. The probe according to claim 6 wherein the quencher is DABCYL.

8. The probe according to claim 4 or claim 5 wherein the chromophore is a fluorophore having an excitation maximum at a wavelength shorter than the 20 excitation maximum of said first fluorophore.

9. The probe of any of claims 4-7 wherein said molecule is a self-hybridizing oligonucleotide having a target complementary region of 7-140 nucleotides flanked by arms hybridizable to one another to form a 25 hybrid 3-25 nucleotides in length.

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10.. The probe according to claim 1 comprising  
two non-FRET pairs of labels.

11. A kit of reagents for an assay comprising  
detection reagents that include a probe according to any  
5 of claims 1-10.

12. The kit according to claim 11 further  
comprising reagents for performing nucleic acid  
amplification.

13. The kit according to claim 12 wherein said  
10 amplification is selected from the group consisting of  
PCR, 3SR, LCR, Q-Beta replication and NASBA.

14. The kit according to claim 13 wherein said  
amplification is PCR.

15. An assay that includes a step of detecting  
15 that comprises contacting at least one probe according to  
any of claims 1-10 with a sample suspected to contain a  
target for said probe and measuring the change in  
fluorescence.

16. The assay according to claim 15 that is a  
20 multiplex assay.

17. The assay according to either of claims 15  
and 16 which includes amplification by the PCR process  
and detection occurs in real time.

18. An assay that includes detecting by  
25 contacting at least one single-stranded hybridization

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probe with a sample suspected of containing a target for said probe, and measuring the absorbance, said probe being capable of hybridizing with a natural oligonucleotide, said probe containing nucleotides 5 selected from the group consisting of natural nucleotides, modified nucleotides and combinations thereof, and said probe containing a pair of chromophores, wherein interaction of the probe with the target causes the probe to change from a first 10 conformation to a second conformation, thereby changing the distance between said chromophores, and wherein in only one conformation do the chromophores touch sufficiently to detectably change absorbance.

19. The assay according to claim 18 wherein 15 said probe is a self-hybridizing oligonucleotide having a target complementary region of 7-140 nucleotides flanked by arms hybridizable to one another to form a hybrid 3-10 nucleotides in length.

## INTERNATIONAL SEARCH REPORT

International application No.

PCT/US97/06532

## A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : C12Q 1/68; C07H 21/04

US CL : 435/6; 536/24.3

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/6; 536/24.3

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

USPATFULL, WPIDS, MEDLINE

search terms: FRET, hybridization, probe?

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	US 5,391,480 A (DAVIS ET AL) 21 February 1995, col. 2, beginning at line 65.	1-19
X ---	US 5,538,848 A (LIVAK ET AL) 23 July 1996, see the abstract and Figure 1.	1, 3, 4, 5, 6, 8,15,18 ----- 11-14
Y	The Stratagene Catalog. 1988, p.39, see the entire document.	11-14
X ---	US 4,766,062 A (DIAMOND ET AL) 23 August 1988, see the entire document, but note especially Figure 3A-3D. This reference was cited in the application.	1,2,15,17,18 ----- 11- 14, 16

Further documents are listed in the continuation of Box C.  See patent family annex.

* Special categories of cited documents:	"T"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
* "A" document defining the general state of the art which is not considered to be of particular relevance	"X"	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
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* "O" document referring to an oral disclosure, use, exhibition or other means		
* "P" document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search	Date of mailing of the international search report
11 JUNE 1997	06 AUG 1997
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231	Authorized officer  ETHAN WHISENANT
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## INTERNATIONAL SEARCH REPORT

International application No.

PCT/US97/06532

## C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 5,607,834 (BAGWELL) 04 March 1997, see the entire document.	1,3,4,5,6, 8,9,15,18
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Y		7,10,11-14, 16,17,19
A	US 4,752,566 A (COLLINS ET AL) 21 June 1988, cited in the application.	1-19
A	US 5,565,322 A (HELLER) 15 October 1996, cited in the application	1-19
A,E	US 5,622,821 A (SELVIN ET AL) 22 April 1997, see the entire document.	1-19
Y	US 5,210,015 A (GELFAND ET AL) 11 May 1993, see the entire document.	17
A	US 4,725,537 A (FRITSCH ET AL) 16 February 1988, cited in the application.	1-19
A	US 4,725,536 A (FRITSCH ET AL) 16 February 1988, cited in the application.	1-19
A	US 5,532,129 A (HELLER) 02 July 1996, see the entire document.	1-19
A	CARDULLO ET AL. Detection of nucleic acid hybridization by nonradiative fluorescence resonance energy transfer. Proc. Natl. Acad. Sci. USA. December 1988, Vol 85, pages 8790-8794 , see the entire document.	1-19
A	SELVIN P.R. Fluorescence Resonance Energy Transfer. Methods in Enzymology. 1995, Vol 246, pages 300-333, see the entire document.	1-19
X	TYAGI ET AL. Molecular Beacons: Probes that fluoresce upon hybridization. Nature Biotechnology. March 1996, Vol 14, pages 303-308, see the entire document.	1-19
A	MATA YOSHI ET AL. Novel fluorogenic substrates for assaying retroviral proteases by resonance energy transfer. Science. 1990, Vol 247, pages 954-958, see the entire document.	1-19

## INTERNATIONAL SEARCH REPORT

International application No.

PCT/US97/06532

## C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	SIXOU ET AL. Intracellular oligonucleotide hybridization detected by fluorescence resonance energy transfer (FRET). Nucleic Acids Research. 1994, Vol 22, No. 4, pages 662-668, see the entire document.	1-19
A	PERKINS ET AL. Fluorescence resonance energy transfer analysis of ribozyme kinetics reveals the mode of action of a facilitator oligonucleotide. Biochemistry. 1996, Vol 35, pages 16370-16377, see the entire document.	1-19
A	EP 0 232 967 A2 (AMOCO CORPORATION) 19 August 1987, see the entire document.	1-19

